

structure than that of the azo form. The creation of this cosphere is favored in water or water-like solvents or in the water-rich compositions of the aqueous binaries. In water-alcohol mixtures, ΔH° becomes negligibly small in compositions where little solvent structure remains and ΔG°_T is governed only by ΔS° .

Whereas the breaks in the plots of ΔG°_T for PND and of \bar{v}_{\max} of TNS against the mole fraction of alcohol must result from changes in the solvent structure, we cannot be certain what that structural change is. The fact that so many properties of these mixtures and of solutes dissolved in them show extrema at 4–6 mol % *tert*-butyl alcohol and 10–15 mol % ethanol has led Franks²¹ and Arnett²² to propose that addition of the alcohols to water leads initially to increased structuring and that the extrema appear at compositions where further addition of alcohol caused breakdown of structure. Yaacobi and Ben-Naim³⁵ interpret the thermodynamics of solution of methane and ethane in ethanol-water mixtures in terms of reinforcement of water structure by ethanol only up to about 3 mol %, with a decrease in water structure between 3 and 20 mol %. By this interpretation, water structure is no longer important above 20 mol %. Proton chemical shift data show that addition of alcohol to water enhances hydrogen-bonded structure between 0 and 5 mol % *tert*-butyl alcohol^{36,37} and 0 and 8 mol % ethanol.³⁸

If it is assumed that alcohol-water mixtures containing more than 20 mol % ethanol and 6 mol % *tert*-butyl alcohol are unstructured, changes in the values of \bar{v}_{\max} for TNS and $\log K_T$ for PND in the unstructured compositions can be used to estimate the importance of solvent changes such as polarity on the probe properties. The excess changes observed in the water-rich compositions then give an estimate of the contribution of structural factors to the total change. Using the values of \bar{v}_{\max} for TNS and 1,7-ANS in ethanol-water

shown in Figure 4 and extrapolating to 0 and 100% ethanol, we estimate contributions of 4.0 and 2.8 kcal/mol, respectively, to the total change in transition energy from solvent effects not related to structuring. The contributions associated with solvent structuring in this medium are 6.3 kcal/mol for TNS and 4.8 kcal/mol for 1,7-ANS. Using eq 6 and 7 to obtain similar extrapolated values of $\log K_T$ of PND in ethanol-water and *tert*-butyl alcohol-water, we estimate contributions to the total change in ΔG°_T of 1.6 kcal/mol from factors related to structuring and 2.7 kcal/mol from factors not related to structuring in ethanol-water mixtures. The corresponding contributions in *tert*-butyl alcohol-water are 2.0 and 1.9 kcal/mol, respectively. The estimates of contributions to the solvent effects due to the existence of solvent structure may be high, since properties of these mixtures that are related to polarity are not linear functions of mole fraction.

The anomalous solvent effect of TFE on both probes shows that in certain favorable cases, a hydrogen-bonding solvent can interact with dye solute by direct hydrogen bonding. The relatively high acidity and low basicity of the fluorinated alcohols reduce their self-association compared to the hydrocarbon alcohols³⁹ and thus increase their tendency to solvate dyes²⁴ and salts²⁵ by hydrogen bonding. The fact that \bar{v}_{\max} values of TNS in the hydrocarbon alcohols are correlated by δ and that the values for these alcohols fall on the same correlation line with aprotic solvents shows that self-association of the hydrocarbon alcohol dominates the solvent effect, and hydrogen-bonded solvent-dye interactions are not very important. The fact that a direct hydrogen-bonded interaction with the probes can shift their properties in the same direction as the indirect solvent-solvent hydrogen-bonding interaction may lead to confusing interpretations and emphasizes the importance of examining a wide range of solvent types and properties. It is most important in interpreting solvent effects to examine solvents in which there are not cross correlations between solvent properties, as is the case in many alcohol-water mixtures.

(39) J. E. Berger, L. R. Dawson, and H. C. Akstran, *J. Phys. Chem.*, **64**, 1458 (1960).

- (35) M. Yaacobi and A. Ben-Naim, *J. Solution Chem.*, **2**, 425 (1973).
 (36) R. G. Anderson and M. C. R. Symons, *Trans. Faraday Soc.*, **65**, 2550 (1969).
 (37) D. N. Glew, H. D. Mak, and N. S. Rath, *Chem. Commun.*, 264 (1968).
 (38) J. R. Kuppers and N. E. Carriker, *J. Magn. Resonance*, **5**, 73 (1971).

Communications to the Editor

Derivation of (+)- and (–)-C₁₇-Juvenile Hormone from Its Racemic Alcohol Derivative via Fungal Metabolism

Sir:

In our metabolic studies with *Helminthosporium sativum*, a racemic mixture of 10,11-epoxyfarnesol has been transformed into (–)-10,11-dihydroxyfarnesoic acid,¹ which was then chemically converted into an enantiomeric pair of C₁₆-juvenile hormone (JH).² Novel trans and cis hydration mechanisms of the racemic

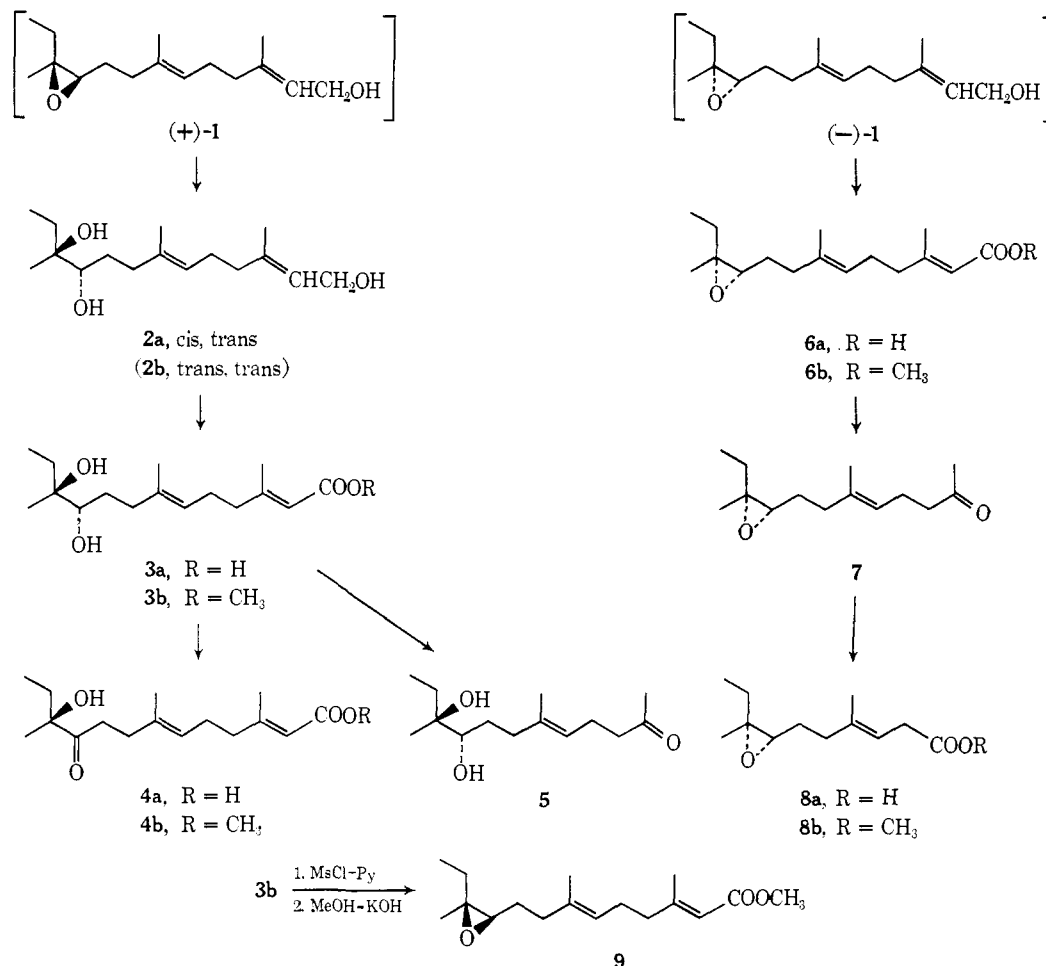
epoxide were also investigated.³ These results can be utilized for optical resolution of racemic epoxy isoprenoids, which is difficult to carry out by the usual resolution methods. In this communication, we describe a derivation of an enantiomeric pair of C₁₇-JH from its racemic alcohol derivative via fungal metabolism. The difference of the metabolic processes between the enantiomers of 10,11-epoxyhomofarnesol is also shown, being compared with those of 10,11-epoxyfarnesol.

Racemic C₁₇-JH has been synthesized in fairly large

- (1) Y. Suzuki and S. Marumo, *Chem. Commun.*, 1199 (1971); *Tetrahedron Lett.*, 1887 (1972).
 (2) Y. Suzuki, K. Imai, S. Marumo, and T. Mitsui, *Agr. Biol. Chem.*, **36**, 1849 (1972).

- (3) Y. Suzuki, K. Imai, and S. Marumo, *J. Amer. Chem. Soc.*, **96**, 3703 (1974).

Scheme I



scale to be used in this work.⁴ The synthetic product contained 23% 2-cis isomer, and, since metabolizing this stereoisomeric mixture with the fungus would lead to contamination of the desired acidic 2-trans metabolites by the corresponding 2-cis ones, the synthetic C_{17} -JH was converted into the alcohol derivative by reduction ($LiAlH_4$) of the ester group. We had established earlier that racemic *cis,trans*-10,11-epoxyfarnesol was metabolized only into $(-)$ -*trans,trans*-10,11-dihydroxyfarnesoic acid and not into the *cis,trans* acid. Thus, the racemic *trans,trans,cis* and *cis,trans,cis* mixture of 10,11-epoxyhomofarnesol (1) was shaken with the precultured mycelia of *Helminthosporium sativum* in the modified Czapek-Dox medium in the same way as in the case of epoxyfarnesol.¹ The extent of the metabolism was checked by tlc, and agitation of the culture was stopped when the substrate disappeared. Chromatographic separation (silica gel) of the ethyl acetate extracts of the culture filtrate gave seven metabolites (Scheme I), *i.e.*, three neutral compounds, 2a (yield 13.5%), 5 (5.9%), and 7 (3.4%), and four acidic ones, 3a (20.6%), 4a (1.3%), 6a (1.9%), and 8a (2.4%), which were purified as their methyl esters.

3b (oil); $C_{17}H_{30}O_4$, m/e 280 ($M^- - H_2O$); $[\alpha]_D -10.2^\circ$ (c 1.1, MeOH); ir (CCl_4) 3500 (OH), 1725 cm^{-1} (ester); nmr ($CDCl_3$) δ 0.94 (t, $J = 7.5$ Hz, $C_{11}-CH_2CH_3$), 1.09 (s, $C_{11}-CH_3$), 1.62 (bs, C_7-CH_3), 1.96 (bs, two OH), 2.15 (bs, C_3-CH_3), 3.38 (dd, $J \approx 2.5, 10.0$ Hz, $C_{10}-CH$), 3.71 (s, $COOCH_3$), 5.17 (bs, C_6-H), 5.70 (bs,

(4) K. Mori, A. Sato, and M. Matsui, *Agr. Biol. Chem.*, **36**, 1931 (1972).

C_2-H). These spectra are unequivocally indicative of **3b** being methyl $(-)$ -*trans,trans*-10,11-dihydroxyhomofarnesoate. Conversion of **3b** into optically active C_{17} -JH was achieved by mesylation (mesyl chloride in pyridine) followed by base treatment (MeOH-KOH). The product (**9**) showed nmr and ir spectra completely identical with those of C_{17} -JH.^{4,5} The optical rotation of **9**, $[\alpha]_D +11.7^\circ$ (c 0.6, MeOH), is quite close to that of (10*R*,11*S*)- C_{18} -JH ($+12.2^\circ$) reported by Loew and Johnson,⁶ indicating that the $(+)$ - C_{17} -JH prepared in the present work must have a fairly high optical purity. This chemical conversion also established the absolute stereochemistry of **3b** to be 10*S*,11*S*.

6b (oil); $C_{17}H_{28}O_3$, m/e 280 (M^+); $[\alpha]_D -12.8^\circ$ (c 0.2, MeOH). The spectral data (nmr and ir) were completely identical with those of C_{17} -JH, showing that the $(-)$ -hormone (as a free acid) has been directly obtained as the result of fungal metabolism. The optical rotation measured was slightly larger than the reported one (-11.7°) of $(-)$ - C_{18} -JH,⁶ suggesting that **6b** must be in a highly pure state. Both enantiomers of C_{17} -JH were thus obtained *via* the fungal metabolic action on the racemic alcohol derivative.⁷

(5) A. S. Meyer, H. A. Schneiderman, E. Hanzmann, and J. H. Ko, *Proc. Nat. Acad. Sci. U. S. A.*, **60**, 853 (1968).

(6) P. Loew and W. S. Johnson, *J. Amer. Chem. Soc.*, **93**, 3765 (1971).

(7) Improvement of the low yield of $(-)$ - C_{17} -JH from the culture filtrate by optimizing the metabolic conditions was not pursued, owing to an insufficient amount of the synthesized substrate. Attempts to derive chemically $(-)$ - C_{17} -JH from methyl $(-)$ -10,11-dihydroxyhomofarnesoate (**3b**) by the procedure carried out successfully in the case of C_{16} -JH (halogenation or mesylation of the C_{11} -hydroxyl, followed by base treatment) have failed.

